

## Hamster CHO cell host protein (CHO-HCP) ELISA Kit

Product Number: ELK00J0023

---

### Shipping and Storage

Store at 2-8°C, away from light and moisture. Validity period: 6 months.

### Component

Component	96-Well Configuration	48-Well Configuration
Microplate (96-well, pre-coated)	12 holes × 8 strips	12 holes × 4 strips
Standards	0.3mL × 6 pipes	0.3mL × 6 pipes
Sample diluent	6mL	3mL
HRP-Conjugated Detection Antibody	10mL	5mL
20× washing buffer	25mL	15mL
TMB Substrate A	6mL	3mL
TMB Substrate B	6mL	3mL
Stop solution	6mL	3mL
Plate Sealer	2 sheets	2 sheets

Note: The concentrations of standard samples (S0-S5) are 0, 15, 30, 60, 120, and 240 ng/mL, respectively.

### Description

The reagent kit adopts a double antibody one-step sandwich enzyme-linked immunosorbent assay (ELISA). Add the sample, standard, and HRP labeled detection antibody to the pre coated micropores of hamster CHO cell host protein (CHO-HCP) antibody, incubate and thoroughly wash. Using substrate TMB for color development, TMB is converted to blue under the catalysis of peroxidase and to the final yellow under the action of acid. The depth of color is positively correlated with the hamster CHO cell host protein (CHO-HCP) in the sample. Measure the absorbance (OD value) at a wavelength of 450nm using an enzyme-linked immunosorbent assay (ELISA) reader and calculate the sample concentration.

### Specimen requirements

1. Serum: Collect with sterile tubes, let the blood naturally coagulate at room temperature for 120 minutes or overnight at 2-8°C, centrifuge at 2-8°C for 20 minutes (3000 rpm), carefully collect the supernatant for testing, or store at -20°C or -80°C for later use. But repeated freezing and thawing should be avoided.
2. Plasma: EDTA or heparin sodium should be selected as anticoagulants according to the requirements of the specimen. After mixing evenly, centrifuge at 2-8°C for 20 minutes (3000 rpm), carefully collect the supernatant for detection, or store at -20°C or -80°C for later use. But repeated freezing and thawing should be avoided.
3. Urine, pleural and peritoneal fluid, cerebrospinal fluid, saliva: collected using sterile tubes, centrifuged at 2-8°C for 20 minutes (3000 rpm). Carefully collect the supernatant or store it at -20°C or -80°C for future use. But repeated freezing and thawing should be avoided.
4. Cell culture supernatant: Collect using sterile tubes, centrifuge at 2-8°C for 20 minutes (3000 rpm), carefully collect the supernatant, or store at -20°C or -80°C for later use. But repeated freezing and thawing should be avoided.
5. Animal tissue samples: Wash the tissue with pre cooled PBS (0.01M, pH=7.4) to remove residual blood, weigh and cut the tissue into small pieces. Mix the shredded tissue with the corresponding volume of PBS (usually in a weight to volume ratio of 1:9, for example, 1g of tissue sample corresponds to 9mL of PBS, the specific volume can be adjusted appropriately according to experimental needs, and records should be kept. Recommend adding protease inhibitors to PBS and grinding thoroughly on ice (at low temperature) in a glass homogenizer. Alternatively, grinding can be performed in a tissue grinder. If further lysis of

**For Research Use Only**

tissue cells is required, the homogenate can be sonicated and finally centrifuged at  $5000 \times g$  for 10 minutes at 2-8°C. The supernatant can be collected for testing. For other tissue samples that are not thoroughly homogenized in a glass homogenizer or tissue grinder, they should be thoroughly ground in liquid nitrogen.

6. Plant tissue samples: Wash the tissue with pre cooled PBS (0.01M, pH=7.4) to remove residual soil, pesticides, etc. After weighing, cut the tissue into small pieces. Mix the shredded tissue with the corresponding volume of PBS (usually in a weight to volume ratio of 1:9, for example, 1g of tissue sample corresponds to 9mL of PBS, the specific volume can be adjusted appropriately according to experimental needs, and records should be kept. Recommend adding protease inhibitors to PBS and grinding thoroughly on ice (at low temperature) in a glass homogenizer. Alternatively, grinding can be performed in a tissue grinder. If further lysis of tissue cells is required, the homogenate can be sonicated and finally centrifuged at  $5000 \times g$  for 10 minutes at 2-8°C. The supernatant can be collected for testing. The supernatant should be divided into single doses and stored at -20°C or -80°C, avoiding repeated freezing and thawing.
7. Animal cells: For adherent cells, gently wash the cells with an appropriate amount of pre cooled PBS and separate them by trypsin digestion. Collect cells by centrifugation at  $1000 \times g$  for 5 minutes (suspended cells can be directly collected by centrifugation). Discard the supernatant and wash the cells three times with cold PBS. Resuspend cells in cold PBS at a concentration of  $1 \times 10^7$  cells/mL. Use an ultrasonic disruptor to thoroughly break down cells, causing them to break down and release intracellular components. Centrifuge at 2-8°C for 20 minutes (3000 rpm), then carefully collect the supernatant for testing.
8. Plant cells: Dilute the cell suspension with PBS at pH 7.2-7.4 to achieve a cell concentration of around 1 million/ml, place it on an ice box, and use an ultrasonic disruptor to thoroughly crush the cells. Centrifuge at 2-8°C for 20 minutes (3000 rpm), carefully collect the supernatant for testing.
9. Throat swab: Add 2ml of PBS with pH 7.2-7.4, dissolve the head of the throat swab, shake well, use tweezers to remove the throat swab and squeeze the liquid dry. Centrifuge at 2-8°C for about 20 minutes (2000-3000 rpm), and carefully collect the supernatant. Pack one portion for testing, freeze the rest for later use. If sediment forms during storage, centrifuge again before loading for testing.

Extract specimens as soon as possible after collection, and conduct experiments as soon as possible after extraction. If the experiment cannot be conducted immediately, the sample should be used within 6 days when it can be stored at 2-8°C. Otherwise, it must be stored at -20°C ( $\leq 1$  month) or -80°C ( $\leq 2$  months), and repeated freezing and thawing should be avoided. If precipitation forms during storage, it should be centrifuged again before sample testing. Samples containing  $\text{NaN}_3$  cannot be detected because  $\text{NaN}_3$  inhibits the activity of horseradish peroxidase (HRP).

### Materials required but not supplied

1. ELISA reader (450nm)
2. High precision sampler and nozzle: 0.5-10 $\mu\text{L}$ , 2-20 $\mu\text{L}$ , 20-200 $\mu\text{L}$ , 200-1000 $\mu\text{L}$
3. 37°C constant temperature box

### Reagent Preparation

20 $\times$  dilution of washing buffer: Dilute distilled water at a ratio of 1:20, which means adding 19 parts of distilled water to 1 part of 20 $\times$  washing buffer.

### Washing method

1. Manual board washing: Shake off the liquid in the holes, fill each hole with washing solution, let it stand for 1 minute, shake off the liquid in the holes, pat dry on absorbent paper, and wash the board 5 times in this way.
2. Automatic washing machine: Inject 350 $\mu\text{L}$  of washing solution into each well, soak for 1 minute, and wash the plate 5 times.

### Protocol

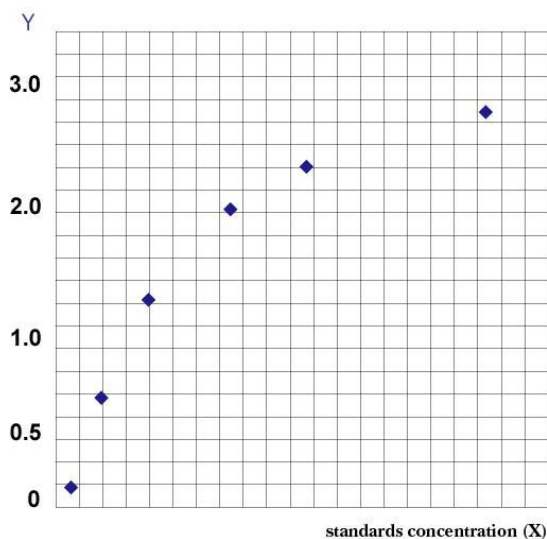
1. Take out the required Flat noodles from the aluminum foil bag after 20 min of room temperature balance, and seal the

remaining Flat noodles with a self sealing bag and put it back at 4°C.

2. Set up standard wells and sample wells, and add 50 $\mu$ L of standard samples of different concentrations to each standard well.
3. Add 10 $\mu$ L of the test sample to the sample well first, and then add 40 $\mu$ L of sample diluent.
4. Add 100 $\mu$ L of horseradish peroxidase (HRP) labeled detection antibody to each well of the standard and sample wells, seal the reaction well with a sealing plate membrane, and incubate at 37°C in a water bath or constant temperature incubator for 60 minutes.
5. Discard the liquid, pat dry on absorbent paper, fill each well with detergent, let it stand for 1 minute, shake off the detergent, pat dry on absorbent paper, repeat washing the board 5 times (or use a board washing machine).
6. Add 50 $\mu$ L of substrate A and B to each well, and incubate at 37°C in the dark for 15 minutes.
7. Add 50 $\mu$ L of termination solution to each well and measure the OD value of each well at a wavelength of 450nm within 15 minutes.

### Result judgment

Draw standard curve: In an Excel worksheet, use the standard concentration as the horizontal axis and the corresponding OD value as the vertical axis to draw a linear regression curve of the standard. Calculate the concentration values of each sample according to the curve equation.



Note: This calibration chart is for reference only. Please refer to the final calibration chart for actual testing.

### Test kit performance

1. Accuracy: The correlation coefficient R value between standard linear regression and expected concentration is greater than or equal to 0.9900.
2. Sensitivity: The minimum detectable concentration is less than 1ng/mL.
3. Detection range: 1-240ng/mL.
4. Specificity: Does not cross react with other soluble structural analogues.
5. Repeatability: The coefficient of variation within and between plates is less than 15%.

### Note

1. The reagent kit should be stored at 2-8°C and equilibrated at room temperature for 20 minutes before use. The concentrated washing solution taken out of the refrigerator will have crystals, which is a normal phenomenon. Heating in a water bath will completely dissolve the crystals before use.
2. The Flat noodles not used in the experiment should be immediately put back into the self sealing bag, sealed (low-temperature drying) and stored.
3. The S0 standard with a concentration of 0 can be considered as a negative control or blank; When operating according to the



## **MEBEP TECH(HK) Co., Limited**

Email: [sales@mebep.com](mailto:sales@mebep.com) Website: [www.mebep.com](http://www.mebep.com)

Tel: +86-755-86134126 WhatsApp/Facebook/Twitter: +86-189-22896756

instructions, the sample has already been diluted 5 times, and the actual concentration of the sample is obtained by multiplying the final result by 5.

4. Strictly follow the time, liquid dosage, and sequence indicated in the instructions for incubation operation.
5. Shake all liquid components thoroughly before use.
6. Take down the enzyme labeled Flat noodles that cannot be used temporarily and put them into aluminum foil bags for standby, and store them at 2-8°C. Suggest using it as soon as possible.

### **Disclaimers**

1. The reagent kit is for research purposes only and should not be used for clinical experiments or mouse experiments. Any consequences arising therefrom shall be borne by the experimenter, and our company shall not be held responsible.
2. Strictly follow the instructions for operation. If the experimenter violates the instructions, the consequences shall be borne by the experimenter.