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Diagnostic Kit for Salmonella DNA

Product Number: DTL0608

Shipping and Storage

1. Store below 30℃. It is valid for 12 months.
2. Transport at normal temperature, not suggested over 14 days.
3. Opened but not completely used the all components should be stored at (-20±5)℃. It is recommended to separate in PCR tubes before refrigeration to avoid repeated freezing and thawing of all reagents next time. It is not recommended to repeat the freeze-thaw cycle more than 7 times.
4. Date of manufacture and term of validity: see the label.

Component

Component	48T
Sal PCR Master Mix	Lyophilized powder ×1 Bottle
Positive Control	100μL
Negative Control	1mL
Redissolved Diluent	1.5mL

Do not mix reagents from different batches.

Description

The kit uses real-time fluorescent PCR detection technology, and adopts a pair of specific primers of Salmonella (Sal) and a specific fluorescent probe to realize qualitative detection of Salmonella pathogens in stool samples. The kit is provided with an Internal Control, which can monitor whether there is PCR inhibitor in the sample to be tested by detecting whether the internal control is normal or not, so as to avoid false negative PCR.

Application

This kit is intended for the in vitro qualitative detection of Salmonella nucleic acids in purulent blood or mucus of feces, and it is suitable for the auxiliary diagnosis of diseases caused by Salmonella. The test results are only for clinical reference and should not be used as the sole criterion for clinical diagnosis.

Applicable instruments

Real-time fluorescence PCR instrument with FAM channel and VIC channel.

Specimen collection

1. Applicable sample type: Feces sample, Suspected contaminated food or water.
2. Sample collection: Feces samples: Collect 0.5-1mL of stool sample, centrifuge at 13000rpm for 2 minutes, and remove the supernatant; Suspected contaminated food sample: The suspected contaminated food is cut by surgery and mixed evenly, take 0.05g and grind it in a grinder.Add 1.5mL of physiological saline and continue grinding. After homogenization, transfer it to a 1.5mL sterilized centrifuge tube, centrifuge at 8000rpm for 2 minutes,collect 100μL of supernatant in 1.5mL sterilized centrifuge tube. Suspected contaminated water: Collect directly 100μL suspected contaminated water for use.
3. Sample storage and transportation: The collection or processing sample should not exceed 24 hours under the conditions of 2℃ ~ 8℃. If long-term preservation is needed, it should be stored below -70℃, and the freezing fusion should not exceed 3 times.

Protocol

1. **Reagent preparation:**

Take out the Sal PCR Master Mix, open the bottle cap according to the arrow direction of the aluminum-plastic cover, add 960μL of Redissolved Diluent, stand for 30 to 60 seconds until the liquid is clear and transparent.

Mix evenly for 15 seconds with a vortex shaker, centrifuge for 3 seconds with a palm centrifuge, and then subpackage it into PCR reaction tubes according to 20μL/ tube.

2. Nucleic acid extraction:

This kit is not included for Nucleic Acid(NA) extraction reagent.

Commercially available extraction kits that have been shown to generate highly purified DNA when following manufacturer's recommended procedures for sample extraction are applicable.

If the extracted DNA is not used immediately, it should be stored below -20℃. For long-term storage, it should be stored below -80℃ and avoid repeated freezing and thawing.

Note: The Negative Control and the Positive Control does not require nucleic acid extraction.

3. Add sample:

The correspond substances were added to that above PCR reaction tubes according to the following table:

Type	Add sample description
Testing Sample	Add 5μL of the extract prepared in step 2 to the reaction tube, and close the tube cover.
Negative Control/ Positive Control	Add 5μL of negative control and positive control to the reaction tube, and cover the tube tightly.

The total reaction volume is 25μL.

After adding the sample, the PCR reaction tubes should be centrifuged for 15s on a palm centrifuge and then delivery to the nucleic acid amplification region. If bubbles are found, the tube wall should be gently flicked to remove bubbles and centrifuged again.

4. PCR amplification:

Place the reaction tube in the automatic fluorescent PCR instrument, set the negative control, positive control, and test sample parameters to perform PCR experiment according to the operating instructions of the instrument, and record the corresponding sample name.

Select FAM channel to detect Sal DNA, select VIC channel to detect the Internal Control. Set the Reaction Volume per Well to 25μL.

(Note: For ABI series instruments, select 'None' under 'Quencher' , and select 'None' as the dye to use as the passive reference.)

Recommended reaction program setting:

Step	Cycles	Temperature	Time	Collect fluorescence signal
1	1 cycle	95℃	2min	No
2	45 cycles	95℃	15sec	No
		60℃	30sec	Yes

5. Result analysis:

After the reaction is completed, the results are automatically saved.

The Start value, End value and Threshold value of the Baseline should be adjusted according to the analyzed image (the user can adjust it according to the actual situation, the Start value can be set at 3~15, the End value can be set at 5~20, the amplification curve of the negative control should be adjusted to be flat or below the threshold line).

Click Analyze for analysis, make the parameters meet the requirements in the following '6.Quality control', and then go to the Plate window to record the Ct value.

6. Quality control

Negative control: FAM and VIC detection channels have no amplification curves.

Positive control: FAM and VIC detection channels have obvious amplification curves, and the Ct value of each channel ≤32.00.

The above requirements must be met at the same time in the same experiment; otherwise this experiment is invalid and needs to

be repeated.

Explanation of Test Result

1. Sal Positive:Ct value ≤40.00 and the curve has a clear index growth curve,IC not required.
2. Sal Negative:Ct value >40.00 or no Ct value, IC Ct value ≤40.00.
3. If the FAM channel amplification curve of the test sample has no logarithmic growth phase and the VIC channel Ct value is >35.00, the test result of the sample is invalid and the cause should be found and eliminated. Collect samples again and repeat the experiment.
4. Note: When testing non-human samples, the VIC channel may not be required.

Limitation

1. Sample detection results are related to sample collection, processing, transportation and preservation quality.
2. If cross-contamination is not controlled during the sample extraction process, false positive results will occur.
3. Positive control and leakage of amplification products can lead to false positive results.
4. The genetic mutations and reorganizations during epidemics can lead to false negative results.
5. Different extraction methods have differences in extraction efficiency, which will lead to false negative results.
6. Reagent transportation, improper preservation, or inaccurate reagent preparation reagent detection performance decreases, and the results of false negative or quantitative detection occur.
7. The results of this test are for reference only. If the diagnosis must be confirmed, please combine clinical symptoms and other test methods.

Performance Parameters

1. Minimum detection limit: The minimum detection limit of this reagent is 500 copies/mL.
2. Precision: The coefficient of variation (CV, %) of the Ct value of the pathogen detection channel is ≤5.00%.
3. Compliance rate of negative/positive reference products: The compliance rate of negative reference products in enterprise reference is 100%, and the compliance rate of positive reference products is 100%.

Note

1. Please read the instructions of this kit carefully before the experiment, and strictly follow the operation steps.
2. Before the test, please be familiar with and master the operation method and precautions of various instruments to be used, and carry out quality control for each experiment.
3. The reaction solution should be stored away from light.
4. Try to avoid bubbles in the reaction, and the tube cover needs to be tight.
5. Use disposable heads, disposable gloves and special work clothes in each district.
6. Sample processing, reagent preparation, and samples need to be performed in different areas to avoid cross -pollution.
7. After the experiment is completed, use 10% hypochlorite or 75% alcohol or ultraviolet light to treat the workbench and pipette.
8. All items in the kit should be treated as pollutants and processed in accordance with the "Biological Safety General of Microbiological Biomedical Laboratory".