MEBEP TECH(HK) Co., Limited
Email: sales@mebep.com Website: www.mebep.com
Tel: +86-755-86134126 WhatsApp/Facebook/Twitter: +86-189-22896756

Diagnostic Kit for Nipah Virus RNA (Real-Time PCRMethod)

Product Number: DTL0162

Shipping and Storage

1. Store below 30°C. It is valid for 12 months.
2. Transport at normal temperature, not suggested over 14 days.
3. Opened but not completely used DGV I/II/III/IV RT-PCR Master Mix and Endogenous Internal Control RT-PCR Master Mix should be stored at (-20±5)°C. It is recommended to separate in PCR tubes before refrigeration to avoid repeated freezing and thawing of all reagents next time. Storage time should not exceed 21 days.
4. Date of manufacture and term of validity: see the label.

Component

Component	48T
NIV RT-PCR Master Mix	Lyophilized powder ×1 Bottle
Positive Control	Lyophilized powder ×1 Tube
Negative Control	1mL
Redissolved Diluent	1.5mL

1. Do not mix reagents from different batches.
2. The reaction system is lyophilized powder that contains all components required for fluorescence PCR, including Taq enzyme, reverse transcriptase, primers, probes, dNTPs, and Mg^{2+}.
3. Add 100μL Redissolved Diluent to the Positive Control, mix well, and use a handheld centrifuge to centrifuge briefly before use.

Description

This kit uses a pair of Nipah virus-specific primers combined with a specific fluorescent probe to perform in vitro amplification and detection of Nipah virus RNA using one-step fluorescent RT-PCR technology, and is used for clinical etiological diagnosis of suspected infected samples.The kit is provided with an Internal Contro（l IC）, which can monitor whether there is PCR inhibitor in the sample to be tested by detecting whether the internal control is normal or not, so as to avoid false negative PCR.

Application

Nipah virus (NiV) is a newly discovered paravirus that is a biosafety level 4 (BSL4) virus. NiV can cause respiratory diseases in pigs and acute febrile encephalitis in humans, and is highly lethal to humans. The mortality rate of humans infected with NiV is as high as 40%-70%, and the possibility of human-to-human transmission was shown in the 2004 Bangladesh outbreak.

This kit is suitable for detecting Nipah virus in samples such as patient blood and throat swabs, and is suitable for auxiliary diagnosis of Nipah virus infection.

Applicable instruments

Real-time fluorescence PCR instrument with FAM,VIC channels.

Specimen collection

1. Applicable sample type: Collect samples such as blood, or throat swabs sample.
2. Sample collection:
   2.1. Blood sample: Blood samples were directly separated into serum or plasma for nucleic acid extraction,If it is plasma, heparin anticoagulation should not be used.
   2.2. Throat swab sample: After collecting the patient's throat swab sample, it can be used directly for nucleic acid extraction.

3. Sample storage and transportation: The collection or processing sample should not exceed 24 hours under the conditions of 2℃ ~ 8℃. If long-term preservation is needed, it should be stored below -70°C, and the freezing fusion should not exceed 3 times.

Protocol

1. Reagent preparation:

Take out the NIV RT-PCR Master Mix, open each bottle cap according to the arrow direction of the aluminum-plastic cover, add 960μL of Redissolved Diluent, strongly mixed on the vortex for more than 1 minute, then stand for 30 ~ 60 seconds until the liquid is clear and transparent. Subpackage it into PCR reaction tubes according to 20μL/ tube.

2. Nucleic acid extraction:

This kit is not included for Nucleic Acid(NA) extraction reagent.

Commercially available extraction kits that have been shown to generate highly purified RNA when following manufacturer's recommended procedures for sample extraction are applicable.

If the extracted RNA is not used immediately, it should be stored below -20℃. For long-term storage, it should be stored below -80℃ and avoid repeated freezing and thawing.

Note:The Negative Control and the Positive Control does not require nucleic acid extraction. The Positive Control needs to be redissolved with 100μL of redissolved diluent and mixed well before use.

3. Add sample:

The correspond substances were added to that above PCR reaction tubes according to the following table:

Type	Add sample description
Testing Sample	Add 5μL of the extract prepared in step 2 to the reaction tube of NIV RT-PCR Master Mix, and close the tube cover.
Negative Control/ Positive Control	Add 5μL of negative control and positive control to the reaction tube, and cover the tube tightly.

The total reaction volume is 25μL.

After adding the sample, the PCR reaction tubes should be mixed well and centrifuged for 5s on a palm centrifuge and then delivery to the nucleic acid amplification region. If bubbles are found, the tube wall should be gently flicked to remove bubbles and centrifuged again.

4. RT-PCR amplification:

Place the reaction tube in the automatic fluorescent PCR instrument, set the negative control, positive control, and test sample parameters to perform RT-PCR experiment according to the operating instructions of the instrument, and record the corresponding sample name.

Select FAM channel to detect Nipah virus nucleic acid, VIC channel to detect Internal Control. Set the sample reaction system to 25μL.

(Note: For ABI series instruments, select 'None' under 'Quencher', and select 'None' as the dye to use as the passive reference.)

Recommended reaction program setting:

Step	Cycles	Temperature	Time	Collect fluorescence signal
1	1 cycle	50℃	10min	No
2	1 cycle	95℃	2min	No
3	45 cycles	95℃	15sec	No
		60℃	30sec	Yes

5. Result analysis:

After the reaction is completed, the results are automatically saved.

The Start value, End value and Threshold value of the Baseline should be adjusted according to the analyzed image (the user can adjust it according to the actual situation, the Start value can be set at 3~15, the End value can be set at 5~20, the amplification curve of the negative control should be adjusted to be flat or below the threshold line).

Click Analyze for analysis, make the parameters meet the requirements in the following 'Quality control', and then go to the Plate window to record the Ct value.

6. Quality control

Negative Control ： FAM,VIC detection channels has no obvious amplification curve.Internal control will not get amplified in the Negative Controls.Ignore the VIC amplification graph if a late signal appears in the Negative control wells.

Positive Control：FAM,VIC detection channel all has an obvious amplification curve, and the Ct value ≤35.00.

The above requirements must be met at the same time in the same experiment, otherwise this experiment is invalid and needs to be repeated.

Explanation of Test Result

Positive: If the Ct value of the detection channel is ≤40.00, and the curve has an obvious exponential growth curve, then the virus type corresponding to the channel is positive, and there is no IC requirement.

Negative: If the Ct value of the sample test result is >40.00 or there is no Ct value, and the Ct value of the IC is ≤40.00, then the virus type corresponding to the channel is negative.

If the FAM channel amplification curve of the test sample has no logarithmic growth phase and the VIC channel Ct value is >40.00, the test result of the sample is invalid and the cause should be found and eliminated. Collect samples again and repeat the experiment.

Note: When testing non-human samples, the VIC channel not be required.

Limitation

1. Sample detection results are related to sample collection, processing, transportation and preservation quality.
2. There is no control of cross -pollution during the sample extraction process, and false positive results will occur.
3. Positive control and leakage of amplification products can lead to false positive results.
4. The genetic mutations and reorganizations during epidemics can lead to false negative results.
5. Different extraction methods have differences in extraction efficiency, which will lead to false negative results.
6. Reagent transportation, improper preservation, or inaccurate reagent preparation reagent detection performance decreases, and the results of false negative or quantitative detection occur.
7. The results of this test are for reference only. If the diagnosis must be confirmed, please combine clinical symptoms and other test methods.

Performance Parameters

1. Limit of Detection: The minimum detection limit of this reagent is 500 copies/mL.
2. Precision: Repeat detection of the enterprise precision reference product 10 times, and the coefficient of variation (CV, %) of the Ct value of the pathogen detection channel is ≤5.00%.
3. Compliance rate of negative/positive reference products: The compliance rate of negative reference products in enterprise reference is 100%, and the compliance rate of positive reference products is 100%.

Note

1. Please read the instructions of this kit carefully before the experiment, and strictly follow the operation steps.
2. Before the test, please be familiar with and master the operation method and precautions of various instruments to be used, and carry out quality control for each experiment.
3. The reaction solution should be stored away from light.
4. Try to avoid bubbles in the reaction, and the tube cover needs to be tight.
5. Use disposable heads, disposable gloves and special work clothes in each district.
6. Sample processing, reagent preparation, and samples need to be performed in different areas to avoid cross -pollution.
7. After the experiment is completed, use 10% hypochloride or 75% alcohol or ultraviolet light to treat the workbench and pipette.
8. All items in the kit should be treated as pollutants and processed in accordance with the "Biological Safety General of Microbiological Biomedical Laboratory".