

# Campylobacter jejuni (CJ) Nucleic Acid Detection Kit (Fluorescent PCR Method)

**Product Number: DTK269**

---

## Shipping and Storage

1.  $-20^{\circ}\text{C}\pm 5^{\circ}\text{C}$ , stored in the dark, transported, and subjected to repeated freeze-thaw cycles no more than 5 times, with a validity period of 12 months.
2. The collected or processed samples should be stored at  $2^{\circ}\text{C}\sim 8^{\circ}\text{C}$  for no more than 24 hours; If long-term storage is required, it should be stored at  $-70^{\circ}\text{C}$  or below, with no more than 3 freeze-thaw cycles.

## Component

Component	50T
CJ reaction solution	$500\mu\text{L}\times 2$
Enzyme solution	$50\mu\text{L}$
CJ positive quality control product	$250\mu\text{L}$
Negative quality control product	$250\mu\text{L}$

**Note: Different batches of reagents cannot be mixed.**

## Description

This kit is designed with specific primers and probes for the conserved region of the gene of *Campylobacter jejuni*. Fluorescence PCR technology is used for in vitro amplification and detection of the nucleic acid of *Campylobacter jejuni*, which is used for pathogenic diagnosis of suspected infectious materials in clinical practice.

## Application

*Campylobacter jejuni* is a subspecies of the genus *Campylobacter*, belonging to the family Molluscaceae. It has been recognized as an important cause of acute diarrhea. It has been proven to be a zoonotic pathogen, and consuming food and drinking water contaminated with this bacterium can cause acute enteritis, posing a threat to human health.

This kit is suitable for detecting *Campylobacter jejuni* in water sample feces, suspected contaminated water, food and other samples, and is used for auxiliary diagnosis of *Campylobacter jejuni* infection.

## Applicable instruments

ABI7500, Agilent MX3000P/3005P, LightCycler, Bio-Rad, Eppendorf and other series of fluorescence quantitative PCR detectors.

## Specimen collection

Take 0.5-1mL of water sample; Take 1g of suspected contaminated food.

## Protocol

### 1. Sample processing (sample processing area)

#### 1.1. Sample pre-processing

Centrifuge the water sample at 13000rpm for 2 minutes to remove the supernatant; Suspected contaminated food was cut and mixed with surgical scissors, and 0.5g was taken and ground in a grinder. 1.5mL of physiological saline was added and continued to grind. After homogenization, the mixture was transferred to a 1.5mL sterile centrifuge tube and centrifuged at 8000rpm for 2 minutes. 100μL of supernatant was taken and placed in a 1.5mL sterile centrifuge tube; Take

**For Research Use Only**

100μL of suspected contaminated water directly.

## 1.2. Nucleic acid extraction

We recommend using our company's nucleic acid extraction or purification reagents (magnetic bead method or centrifugal column method) for nucleic acid extraction. Please follow the reagent instructions for operation.

## 2. Reagent preparation (reagent preparation area)

Based on the total number of samples to be tested, the required number of PCR reaction tubes is N (N=number of samples+1 negative control tube+1 positive control tube); For every 10 samples, an additional 1 sample is prepared. The preparation of each test reaction system is shown in the table below:

reagent	CJ reaction solution	Enzyme solution
Dosage (sample size N)	19μL	1μL

Transfer the mixed test reaction solution into a PCR reaction tube at a concentration of 20uL per tube.

## 3. Sample addition (sample processing area)

Take 5μL of the nucleic acid, positive control sample, and negative control sample extracted in step 1, and add them to the corresponding reaction tubes. Cover the tubes, mix well, and briefly centrifuge.

## 4. PCR amplification (nucleic acid amplification zone)

4.1. Place the reaction tube to be tested in the reaction tank of the fluorescence quantitative PCR instrument;

4.2. Set the channel and sample information, and set the reaction system to 25μL;

Fluorescence channel selection: Detection channel (Reporter Dye) FAM, Quencher Dye NONE, please do not select ROX reference fluorescence for ABI series instruments, select None.

4.3. Recommended loop parameter settings:

step	Cycles	Temperature	Time	Collect fluorescence signals
1	1 cycle	95°C	2min	No
2	45 cycles	95°C	15sec	No
		60°C	30sec	Yes

## 5. Result analysis and judgment

### 5.1. Result Analysis Condition Setting

(Please refer to the user manuals of each instrument for setting up, taking the ABI7500 instrument as an example)

After the reaction is complete, the results will be automatically saved. Based on the analyzed image, adjust the Start value, End value, and Threshold value of the baseline (users can adjust them according to their actual situation, with Start value set between 3-15 and End value set between 5-20, so that the threshold line is in the exponential period of the amplification curve, and the amplification curve of negative quality control products is flat or below the threshold line). Click Analyze to automatically obtain the analysis results.

### 5.2. Result judgment

Positive: The Ct value of the detection channel is  $\leq 40$ , and the curve shows a significant exponential growth curve;

Negative: The sample test result shows no Ct value and no specific amplification curve.

Suspicious: If the sample test result is  $40 < \text{Ct value} \leq 45$ , it is recommended to repeat the test. If the detection channel is still  $40 < \text{Ct value} \leq 45$  and the curve has a clear exponential growth curve, it is judged as positive. Otherwise, it is judged as negative.

## Quality control standards

Negative quality control product: no specific amplification curve or Ct value display;

Positive quality control product: The amplification curve shows a significant exponential growth period, and the Ct value is  $\leq$

32;

The above conditions should be met simultaneously, otherwise the experiment will be considered invalid.

## Limitations of detection methods

1. The results of sample testing are related to the quality of sample collection, processing, transportation, and preservation;
2. Failure to control cross contamination during sample extraction can result in false positive results;

**For Research Use Only**

3. Leakage of positive controls and amplification products can lead to false positive results;
4. Genetic mutations and recombination of pathogens during epidemics can lead to false negative results;
5. Different extraction methods have differences in extraction efficiency, which can lead to false negative results;
6. Improper transportation, storage, or preparation of reagents can lead to a decrease in reagent detection efficiency, resulting in false negatives or inaccurate quantitative testing results;
7. The test results are for reference only. If a diagnosis is required, please combine clinical symptoms and other testing methods.

**Note**

1. All operations must be strictly carried out in accordance with the instructions;
2. The various components in the reagent kit should be naturally melted, completely mixed, and briefly centrifuged before use;
3. The reaction solution should be stored away from light;
4. Try to avoid the presence of bubbles during the reaction, and cover the tube tightly;
5. Use disposable suction tips, disposable gloves, and specialized work clothes for each area;
6. Sample processing, reagent preparation, and sample addition should be carried out in different areas to avoid cross contamination;
7. After the experiment is completed, treat the workbench and pipette with 10% hypochlorous acid, 75% alcohol, or a UV lamp;
8. All items in the reagent box should be treated as contaminants and disposed of in accordance with the "Biosafety Guidelines for Microbial Biomedical Laboratories".